

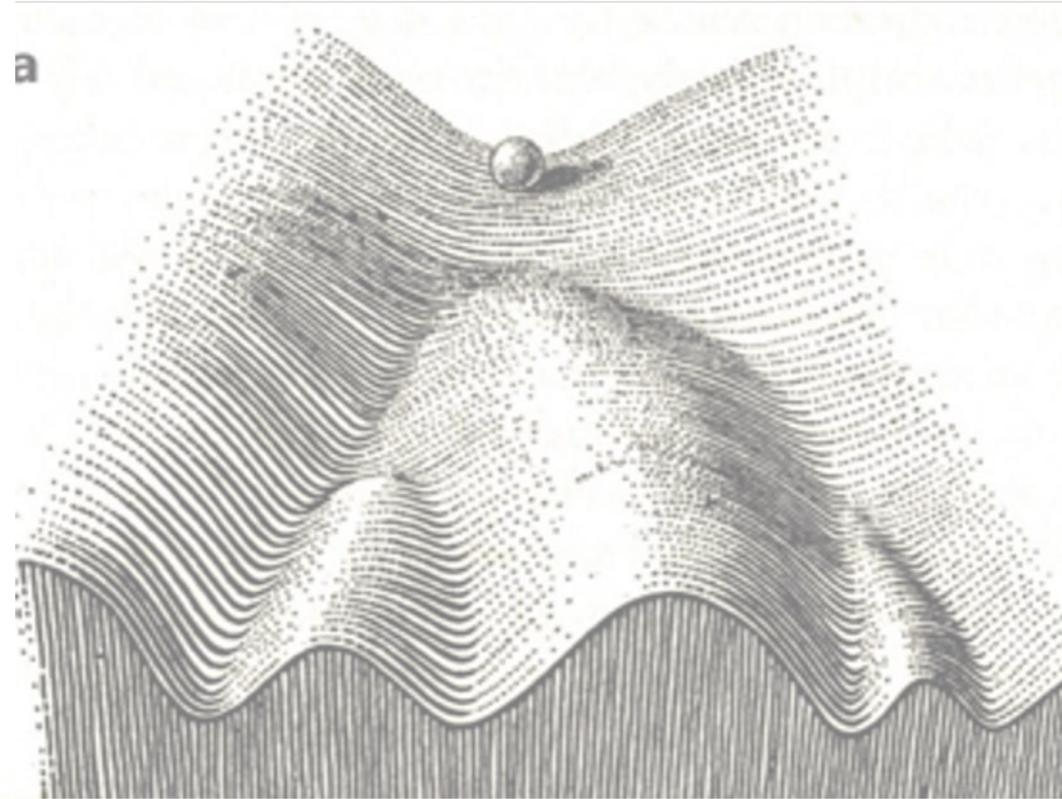
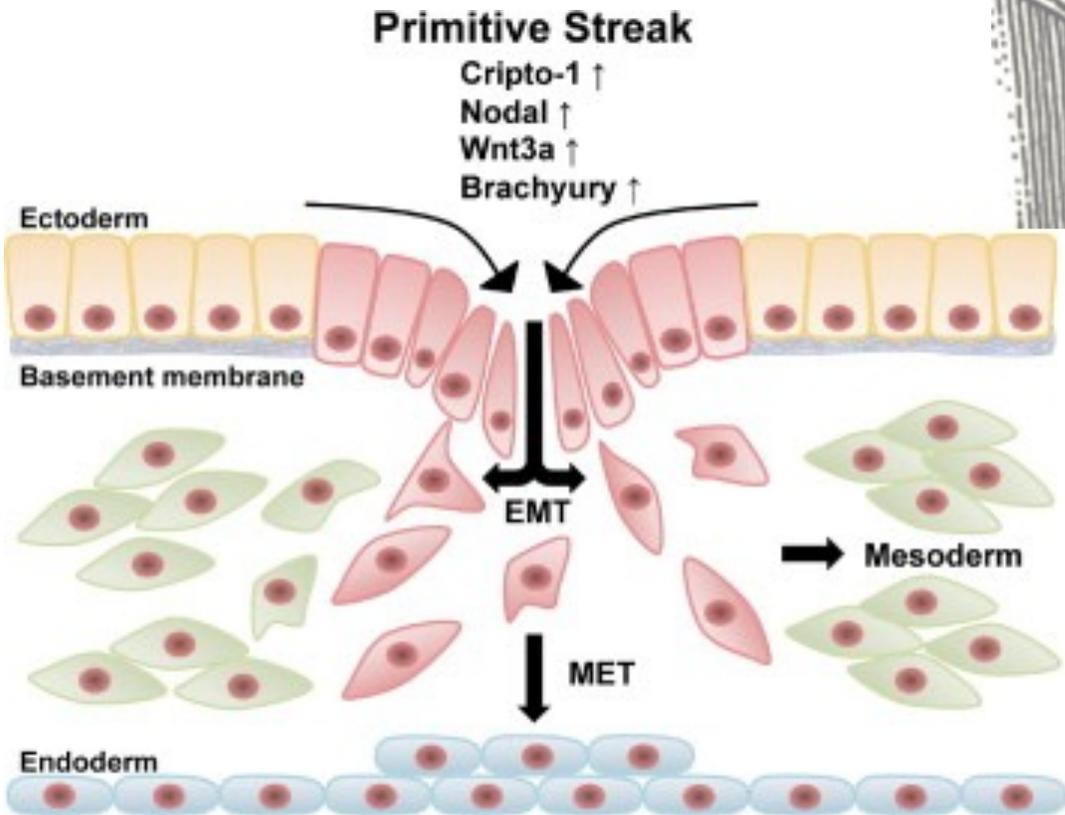
Generalizing RNA velocity to transient cell states through dynamical modeling

Bergen, V., Lange, M., Peidli, S. et al.

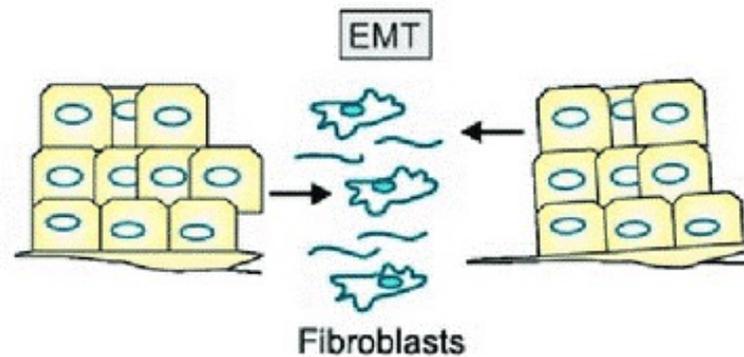
Presenter: Yiyu Pang

Cell fate decision

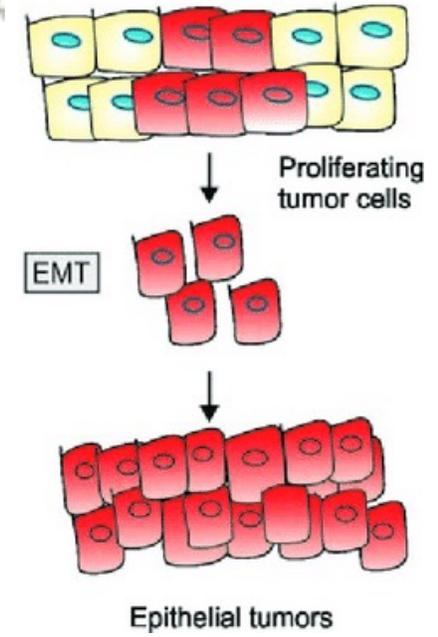
Embryonic development



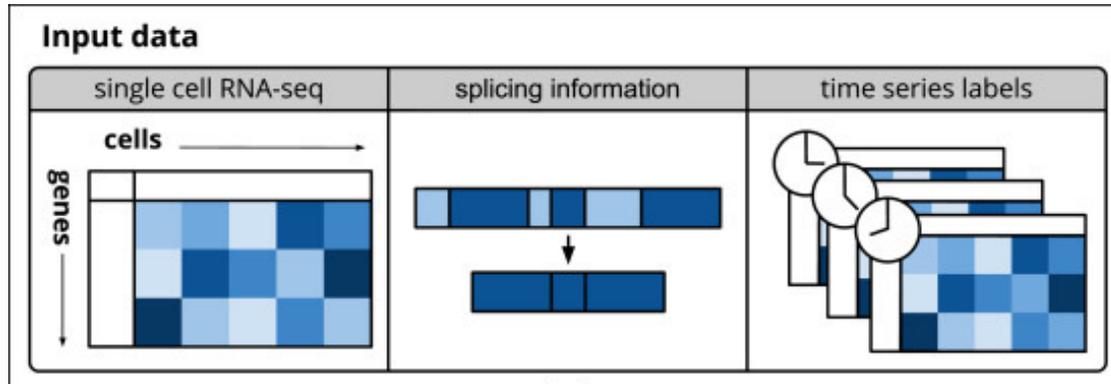
Tissue regeneration



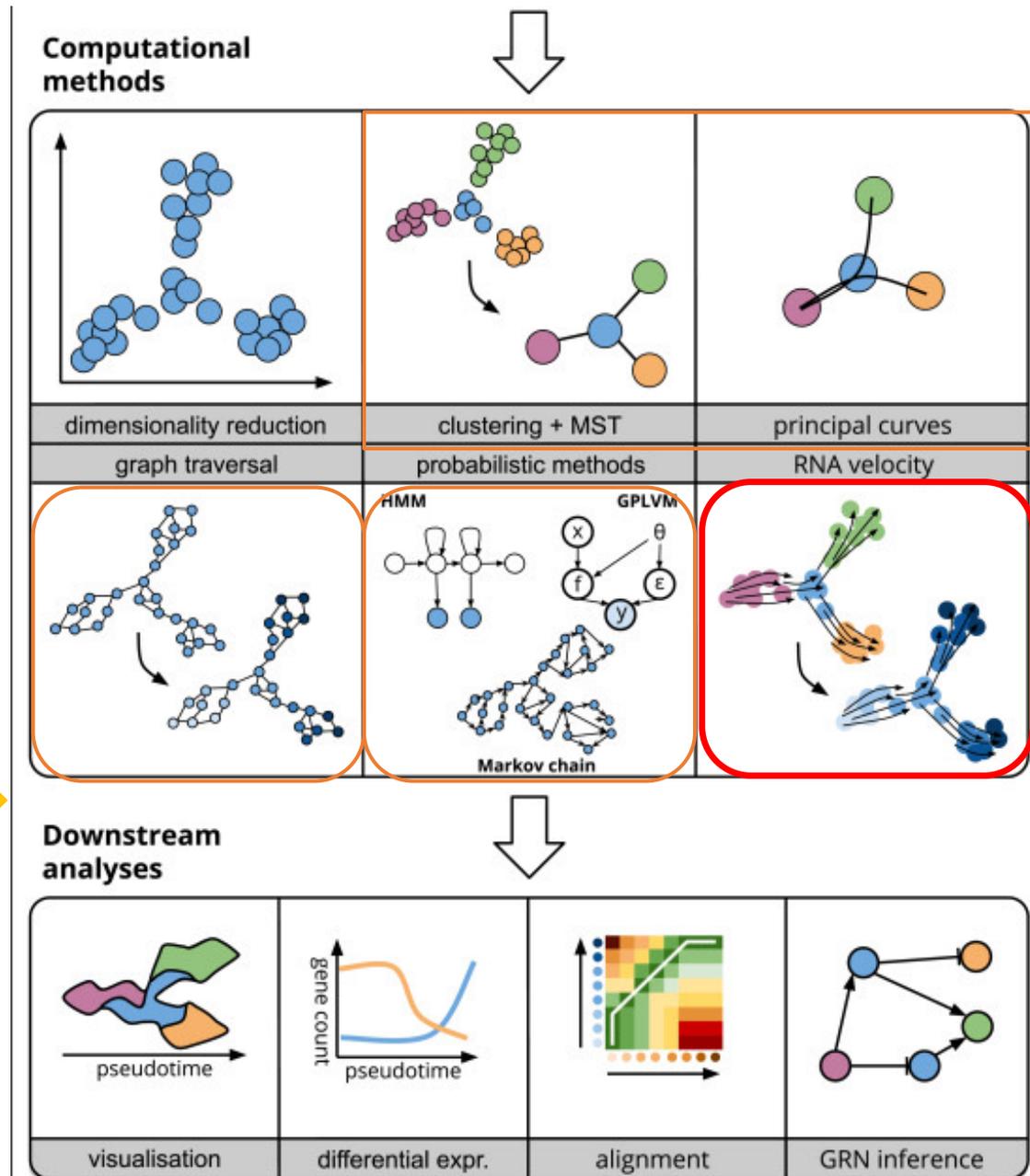
Cancer metastasis



Cell trajectory inference



- MST: minimum weight spanning tree
- Principal curves: smooth one-dimensional curves
 - pass through the middle of a p -dimensional data set
 - providing a nonlinear summary of the data
- Graph traversal: the process of visiting (checking and/or updating) each vertex in a graph
 - Depth-first search
 - Breadth-first search
- Graph decomposition
- RNA velocity: an indicator of the future state of the cell



The dynamics and regulators of cell fate decisions revealed by pseudotemporal ordering of single cells

Cole Trapnell, Davide Cacchiarelli, Jonna Grimsby, Prapti Pokharel, Shuqiang Li, Michaela Mikkelsen, Kenneth J Livak, Tarjei S Mikkelsen & John L Rinn 

Nature Biotechnology **32**, 381–386 (2014) | [Cite this article](#)

Diffusion pseudotime robustly reconstructs lineage branching

Laleh Haghverdi, Maren Büttner, F Alexander Wolf, Florian Buettner & Fabian J Theis 

RNA velocity of single cells

Gioele La Manno, Ruslan Soldatov, Amit Zeisel, Emelie Braun, Hannah Lidschreiber, Maria E. Kastri, Peter Lönnerberg, Alessandro Furlan, Jean David van Bruggen, Jimin Guo, Xiaoling He, Roger Barker, Erik Sundström, Cramer, Igor Adameyko, Sten Linnarsson  & Peter V. Kharchenko 

Nature **560**, 494–498 (2018) | [Cite this article](#)

137k Accesses | **625** Citations | **664** Altmetric | [Metrics](#)

PAGA: graph abstraction reconciles clustering with trajectory inference through a topology preserving map of single cells

Lauren A. Hamey, Mireya Plass, Jordi Solana, Joakim S. Dahlin, Berthold Göttgens, Fabian J. Theis & Fabian J. Theis 

Article number: 59 (2019) | [Cite this article](#)

136 Citations | **136** Altmetric | [Metrics](#)

Slingshot: cell lineage and pseudotime inference for single-cell transcriptomics

Kelly Street, Davide Risso, Russell B. Fletcher, Diya Das, John Ngai, Nir Yosef, Elizabeth Purdom & Sandrine Dudoit 

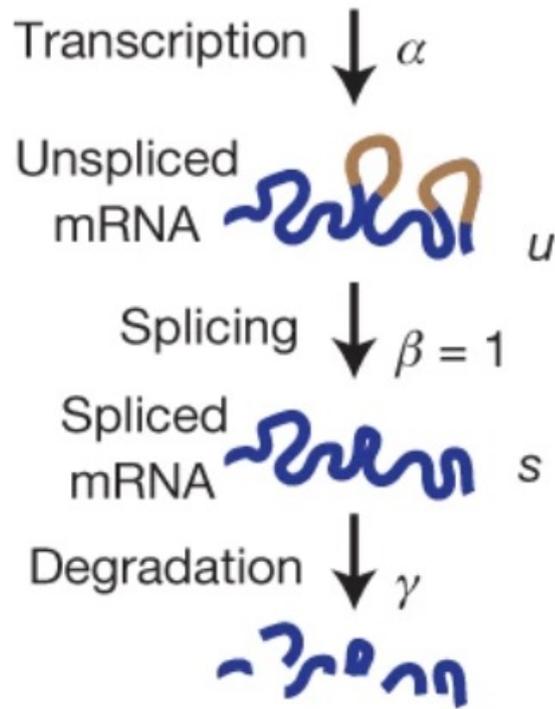
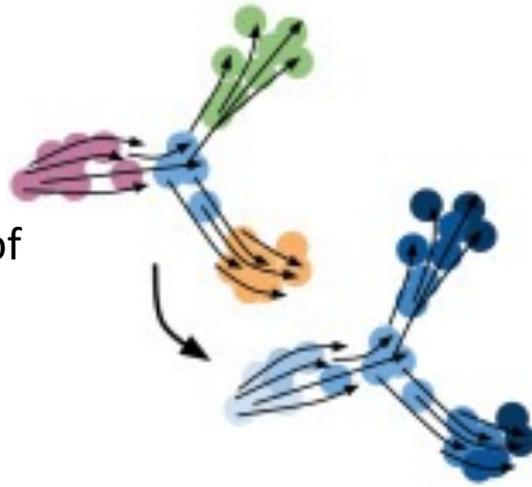
BMC Genomics **19**, Article number: 477 (2018) | [Cite this article](#)

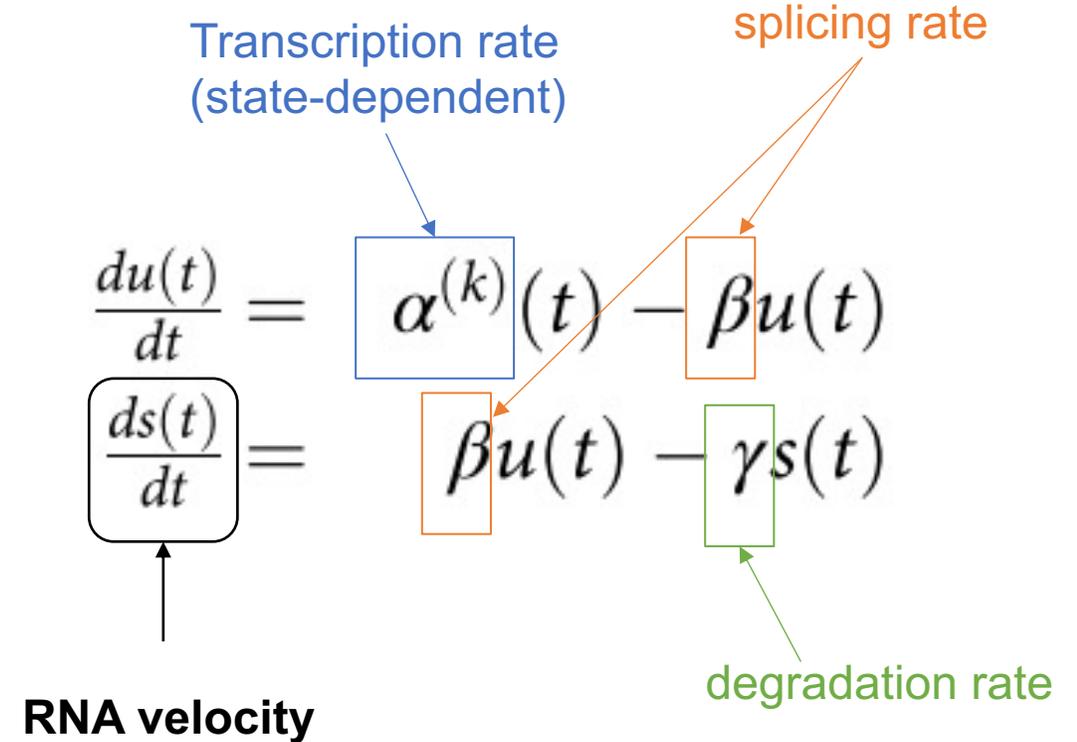
27k Accesses | **296** Citations | **20** Altmetric | [Metrics](#)

RNA velocity

An indicator for

- Future state of the abundance of mature mRNA
- Future state of the cell




$$\frac{du(t)}{dt} = \alpha^{(k)}(t) - \beta u(t)$$
$$\frac{ds(t)}{dt} = \beta u(t) - \gamma s(t)$$

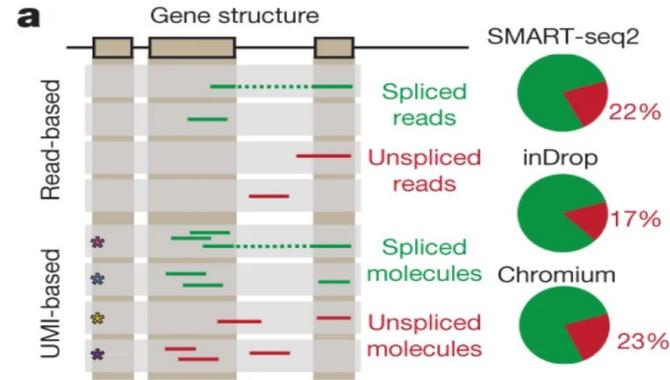
RNA velocity

- u – unspliced /intronic/pre-mRNA abundance
- s – spliced/exonic/mature mRNA abundance

How to estimate RNA velocity?

- Estimate RNA reads
- Examine all data
 - Induction and repression
- Find steady state
- Estimate RNA velocity

Estimate unspliced and spliced reads



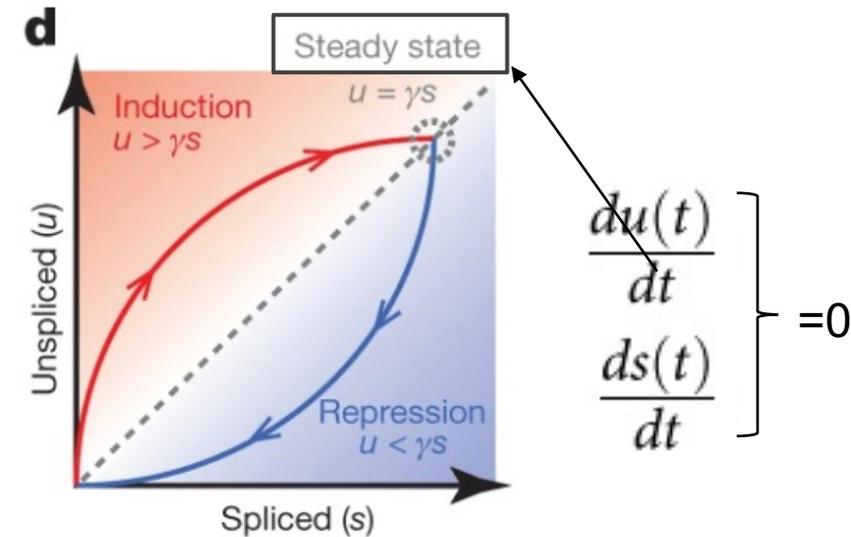
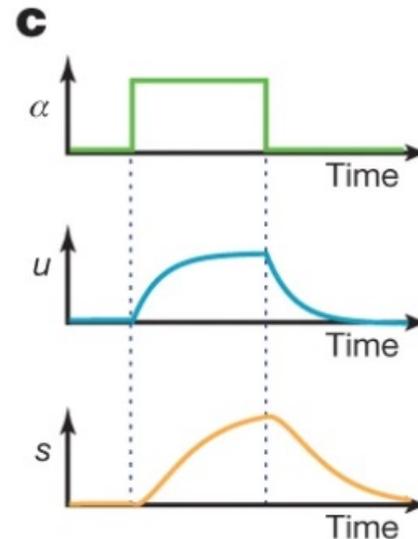
- UMI : unique molecular identifier
- single-cell sequencing protocol:
 - SMART-seq2
 - inDrop
 - Chromium

Transcription rate (state-dependent)

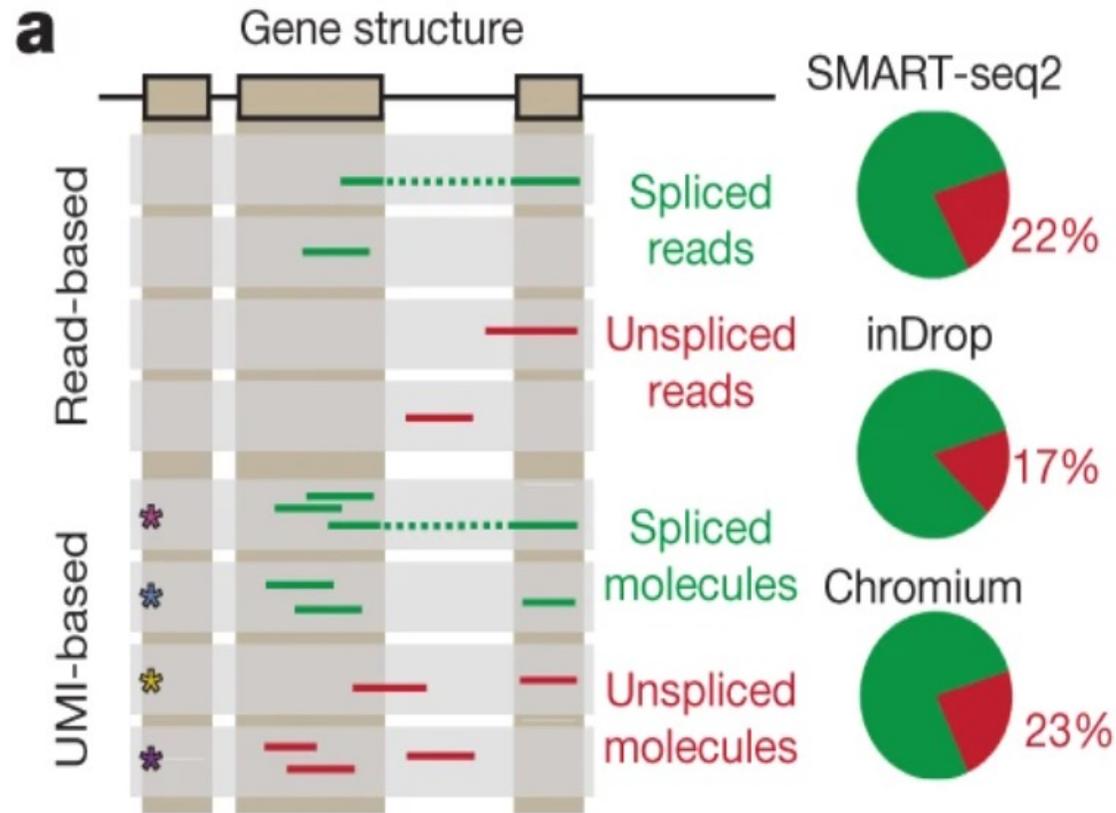
$$\frac{du(t)}{dt} = \alpha^{(k)}(t) - \beta u(t)$$

$$\frac{ds(t)}{dt} = \beta u(t) - \gamma s(t)$$

β assumed to be 1



Estimate unspliced and spliced reads



UMI : unique molecular identifier

- single-cell sequencing protocol:
 - SMART-seq2
 - inDrop
 - Chromium

Estimate RNA velocity

- Governed by two equations

- Solution to equations
- Time course variation of u and s

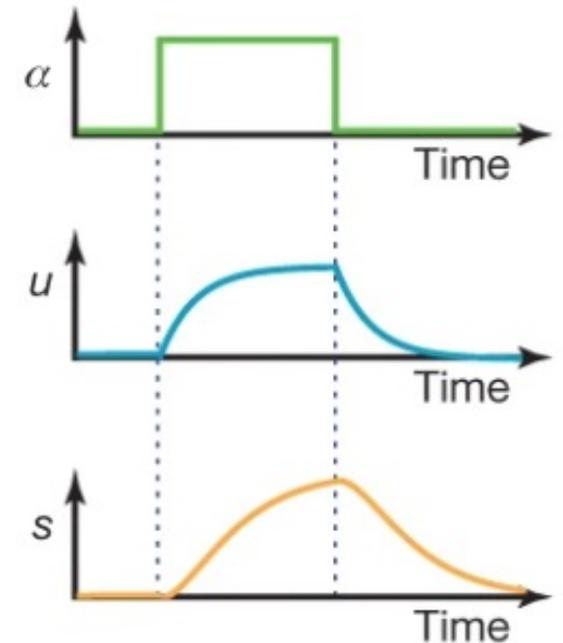
Transcription rate
(state-dependent)

β assumed to be 1

RNA velocity

$$\frac{du(t)}{dt} = \alpha^{(k)}(t) - \beta u(t)$$
$$\frac{ds(t)}{dt} = \beta u(t) - \gamma s(t)$$

degradation rate
Needs to be estimated



- Experimental data:
 - (u, s) from many measurements shown in one plot

- Data fall into two regimes:

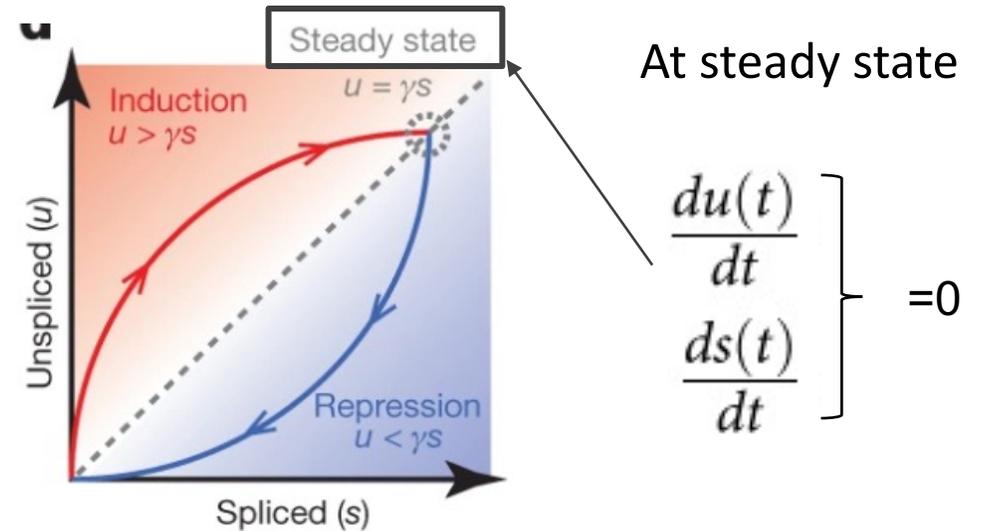
$$\frac{ds(t)}{dt} = \beta u(t) - \gamma s(t)$$

- Up-regulation/induction: $ds/dt > 0$

- Down-regulation/repression: < 0

- Estimate degradation rate

- Using data near steady state



Dashed Line:

When no net change in s

Data near dashed line:

At steady state

Estimate γ using extreme quantile fit

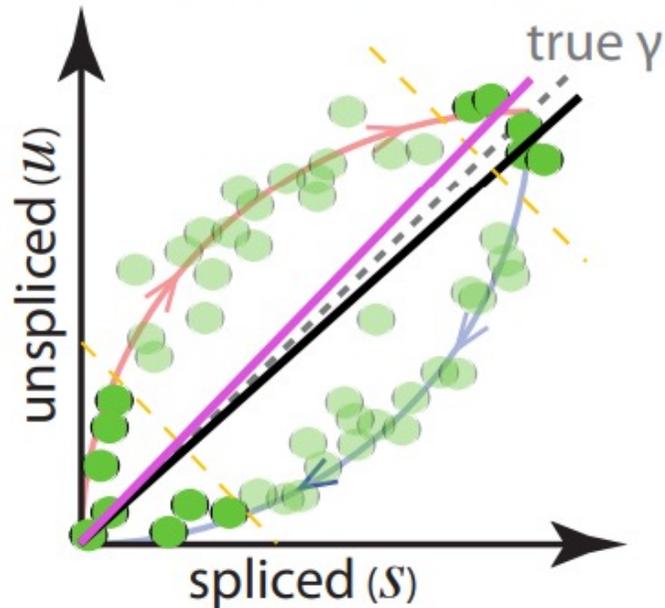
$$v = \frac{ds}{dt} = u(t) - \gamma s(t)$$

Why extreme quantile fit will work?

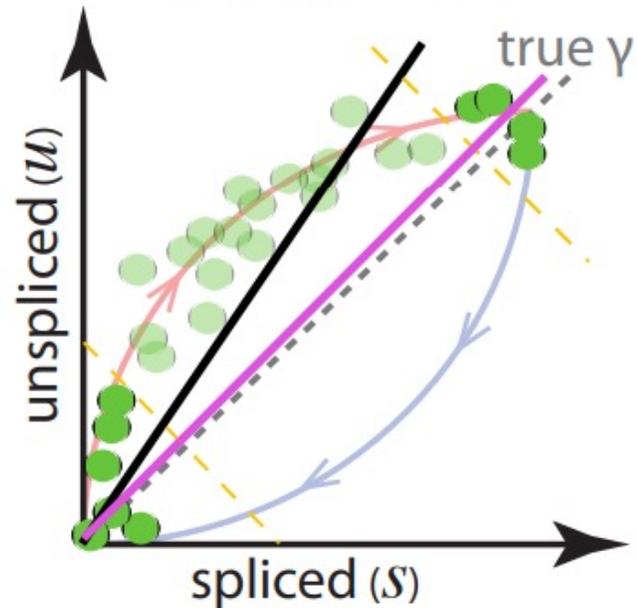
Assume only partial gene regulation is observed

- Only has upregulation observation

a full observation



b upregulation



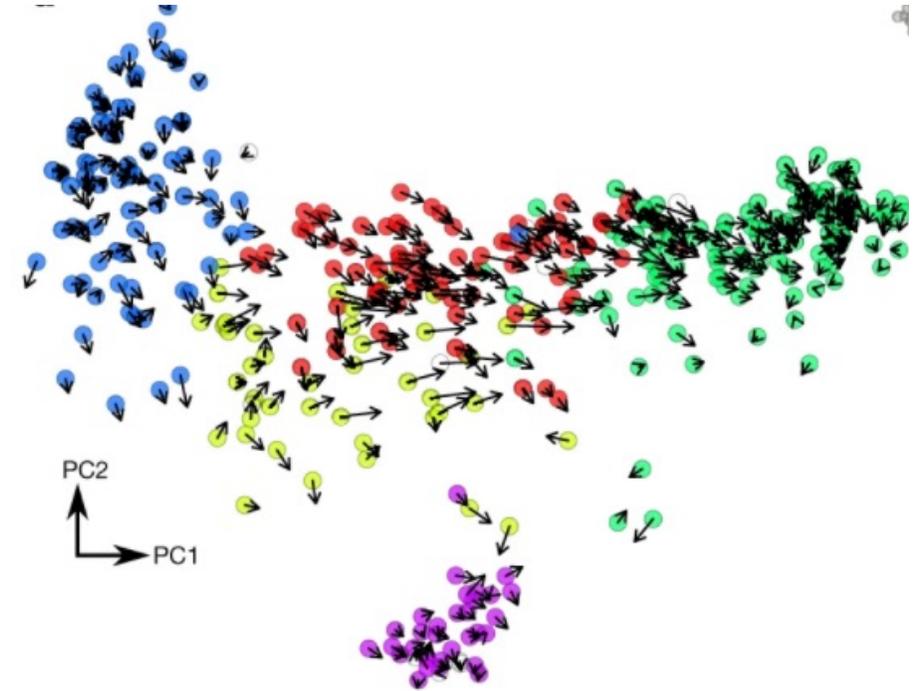
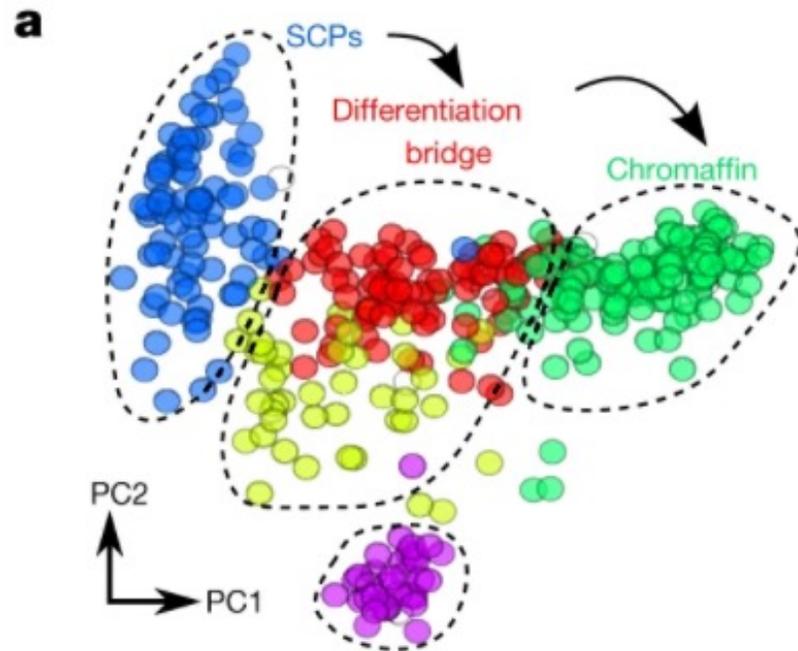
- extreme quantile cell
- other cell
- - true slope
- regular fit
- quantile fit

Extreme quantile fit:

- including only cells near the origin and cells near the upper-right corner of the phase portrait
- Least square fit

Results: Mouse chromaffin cell differentiation

- SCPs: Schwann cell precursors
 - peripheral glial stem cells
- Chromaffin cells
 - neuroendocrine cells found in adrenal medulla
 - constitute the main hormonal component of the autonomic nervous system
 - the principal source for release of catecholamines, including adrenaline, in the systemic circulation
- sympathoblasts



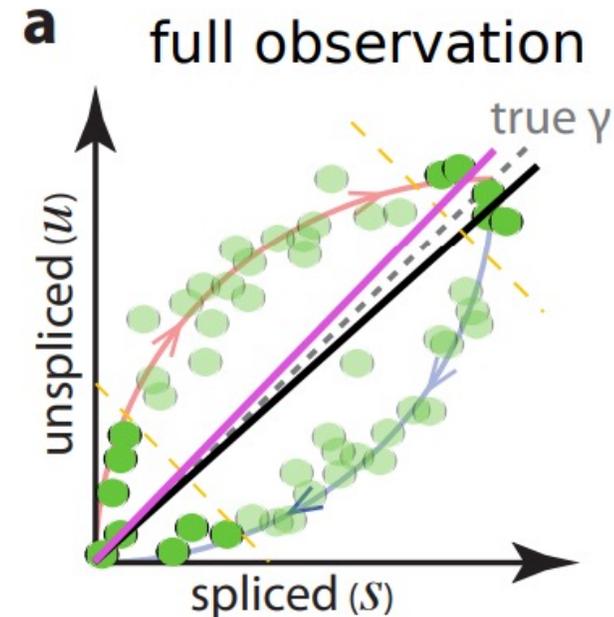
Velocity arrow

- direction
 - expression difference best correlated with the estimated velocity vector
- Length
 - Longer for higher transition probability to other cells

Limitation of the method

- A common splicing rate is assumed across different genes
- Assume the full splicing dynamics are measured
 - Transcriptional induction, repression and steady-state mRNA levels
 - Not realistic

$$\frac{du(t)}{dt} = \alpha^{(k)}(t) - \beta u(t)$$
$$\frac{ds(t)}{dt} = \beta u(t) - \gamma s(t)$$



NATURE BIOTECHNOLOGY

Article | Published: 03 August 2020

Generalizing RNA velocity to transient cell states through dynamical modeling

Volker Bergen, Marius Lange, Stefan Peidli, F. Alexander Wolf  & Fabian J. Theis 

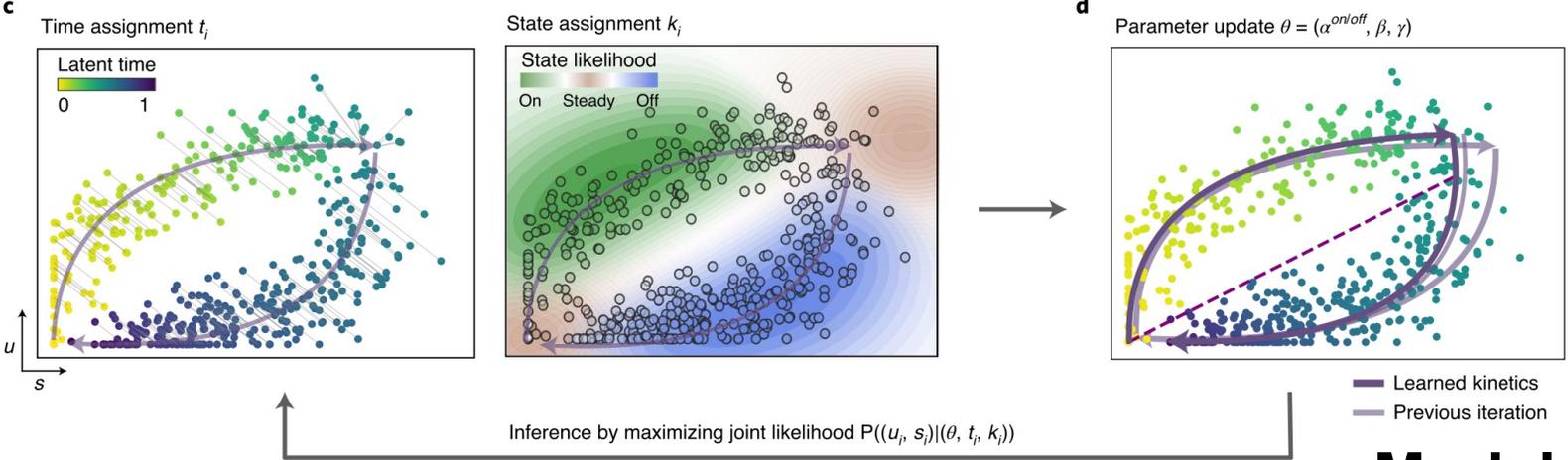
Nature Biotechnology **38**, 1408–1414 (2020) | [Cite this article](#)

33k Accesses | **133** Citations | **322** Altmetric | [Metrics](#)

Generalized RNA velocity

Latent time t_i

- Induction and repression process: a continuous time process
- the latent time: from the starting point right before the induction
- Cell at different latent time with (u, s) changes



β not 1 anymore

$$\frac{du(t)}{dt} = \alpha^{(k)}(t) - \beta u(t)$$

RNA velocity $\frac{ds(t)}{dt} = \beta u(t) - \gamma s(t)$

Model parameters: $\theta = (\alpha^{(k)}, \beta, \gamma)$

Different regimes:

- 1: up-regulation
- 0: down-regulation
- SS_1 : active steady state
- SS_0 : inactive steady state

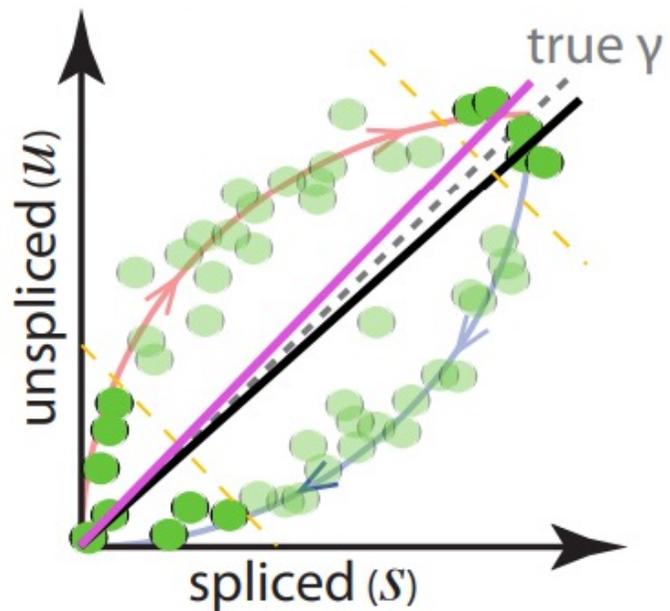
$$k_i \in \{1, 0, SS_1, SS_0\}$$

Model of continuous time process to estimate

- Model parameters: α, β, γ
- Latent time for each cell

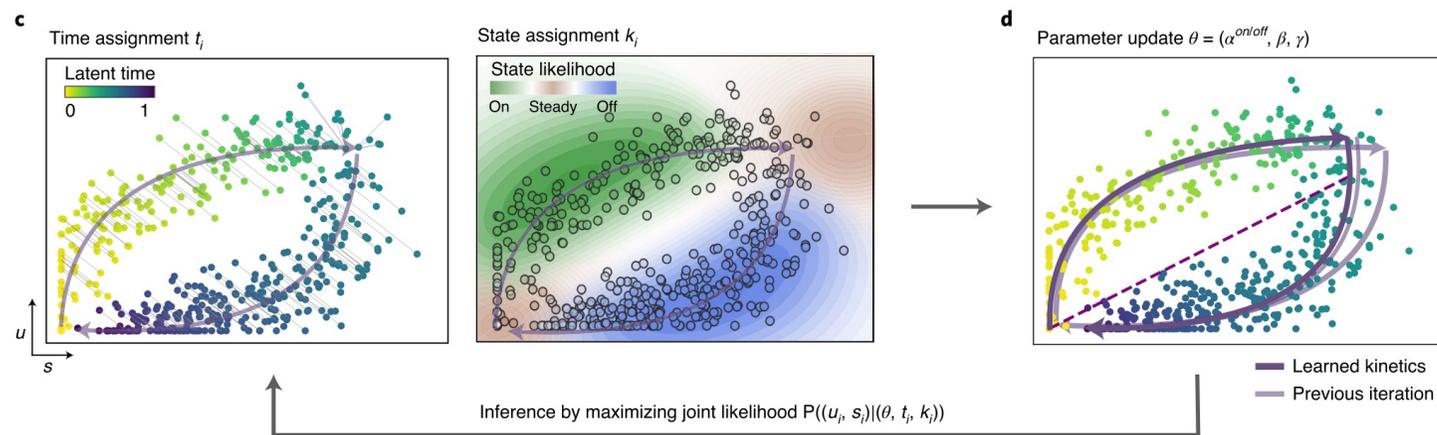
Results: best fit summarized as gray curves.

Steady-state model



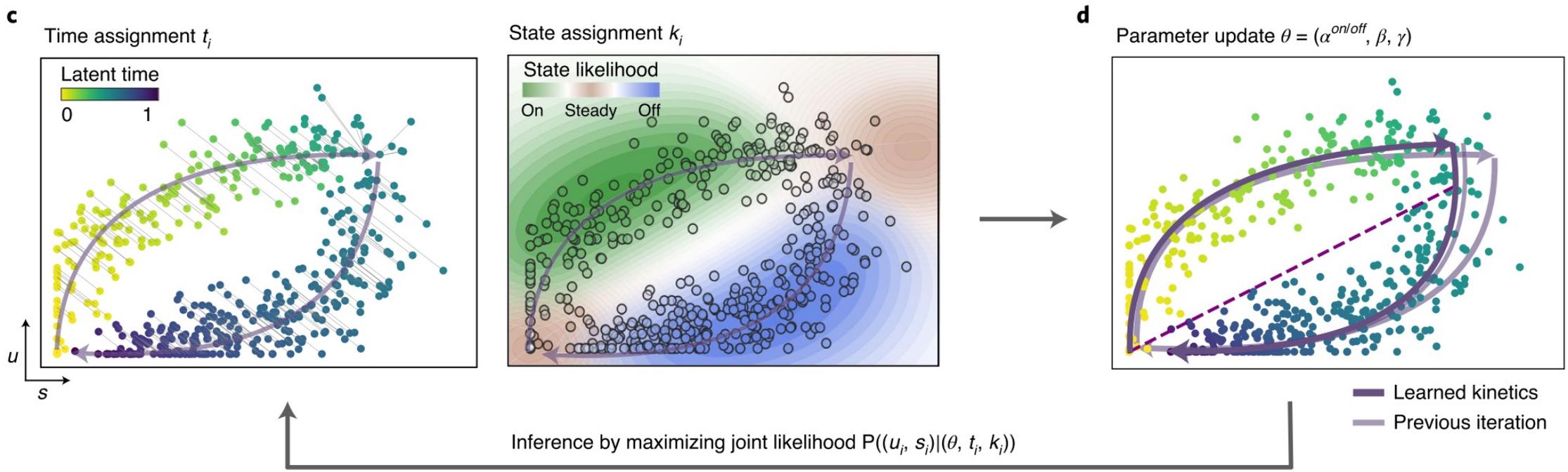
Paper 1

Dynamical model



Paper 2

Generalized RNA velocity



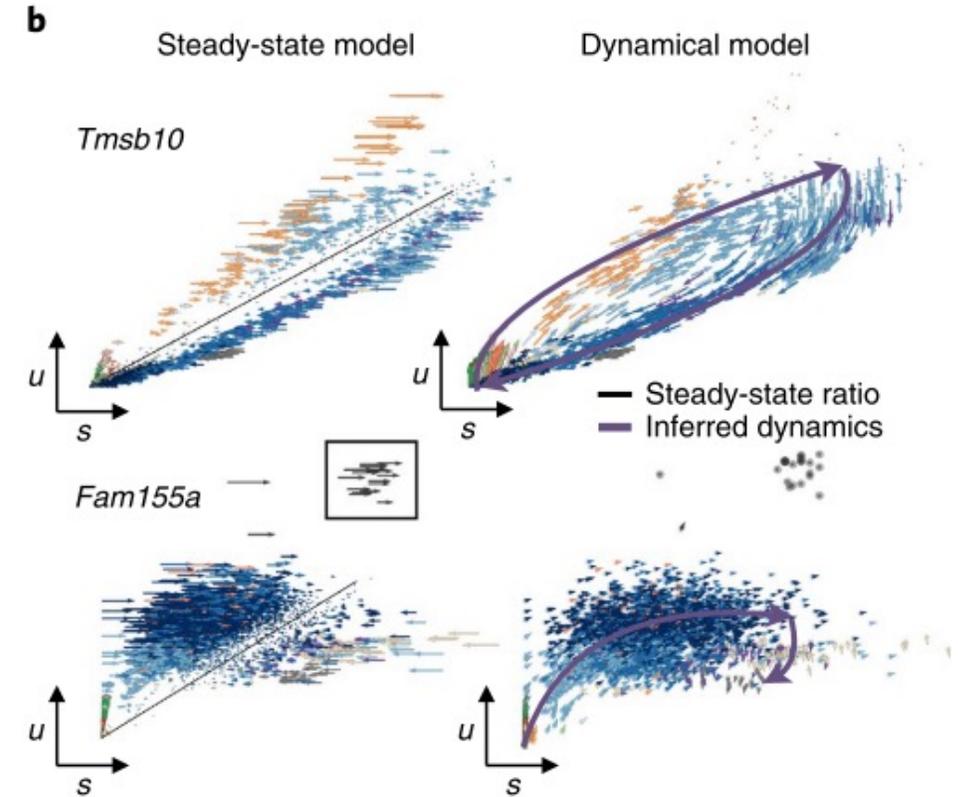
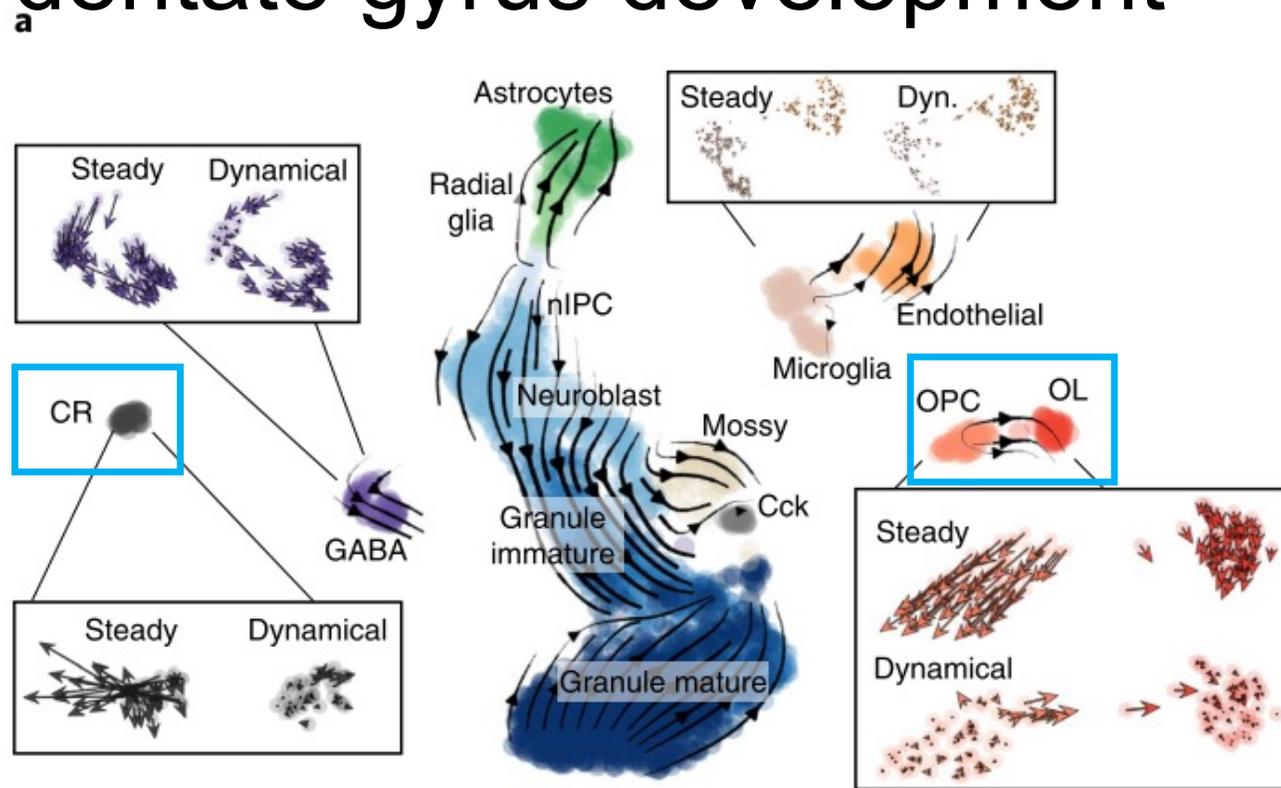
find a phase trajectory specified by $\hat{x}(t)$ that best describes the observations.

model estimate be $\hat{x}(t) = (\hat{u}(t), \hat{s}(t))$

observations u_i^{obs} and s_i^{obs}

The likelihood for a particular gene $\mathcal{L}(\theta) = \frac{1}{\sqrt{2\pi}\sigma} \exp\left(-\frac{1}{2n} \sum_i \frac{\|x_i^{obs} - x_{t_i}(\theta)\|^2}{\sigma^2}\right)$.

Results: Resolving the heterogeneous population kinetics in dentate gyrus development

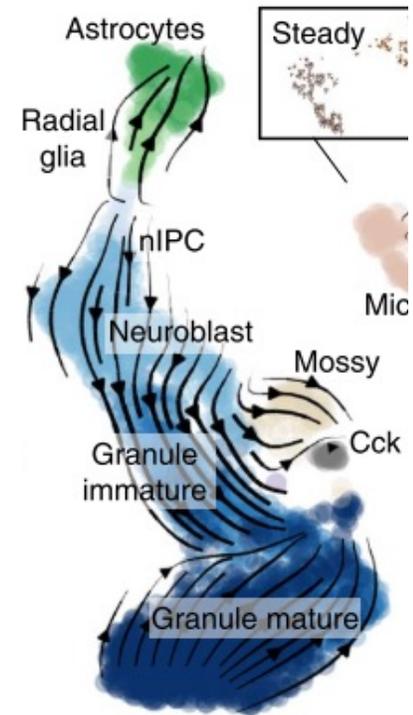
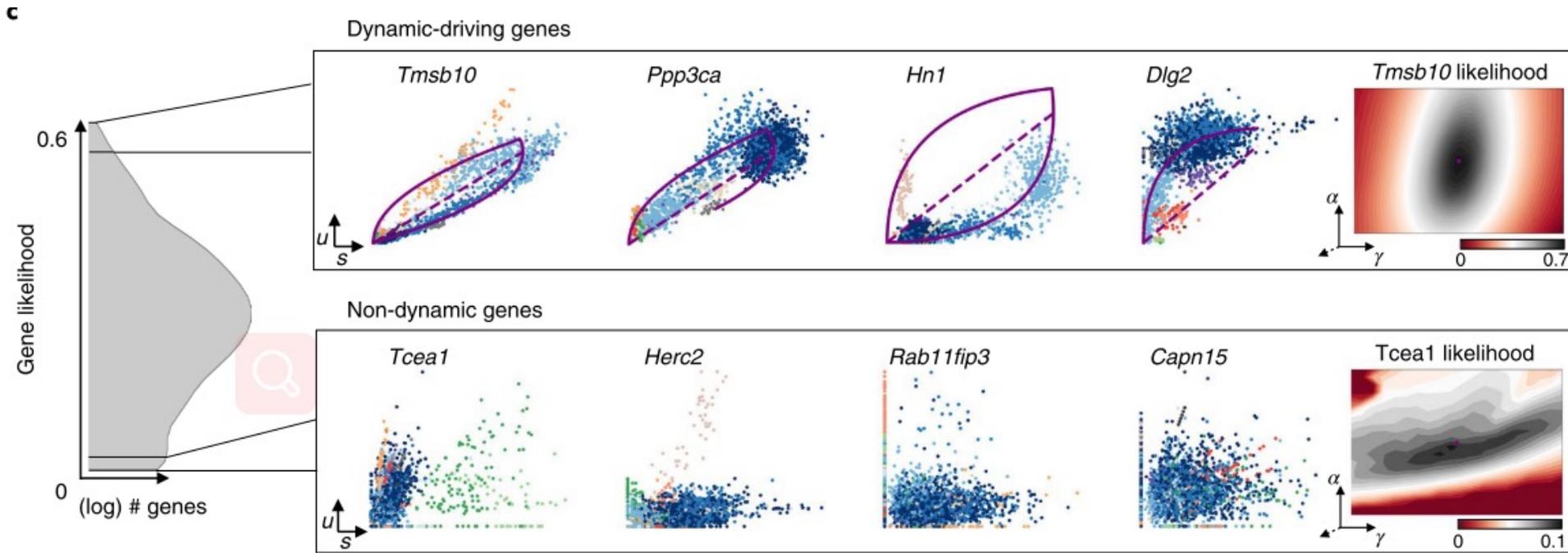


- Only **dynamical model** correctly identifies
 - the OPCs differentiating into OLs
 - CR cells as terminal.

- Velocity of CR cells
 - Steady-state model
 - erroneously high velocities to CR cells
 - *Fam155a* phase portrait
 - while expression patterns show no evidence for any further maturation within the CR population.

OPCs : oligodendrocyte precursor cells
 OLs: myelinating oligodendrocytes
 CRs: Cajal–Retzius cells

Results: Identify putative driver genes



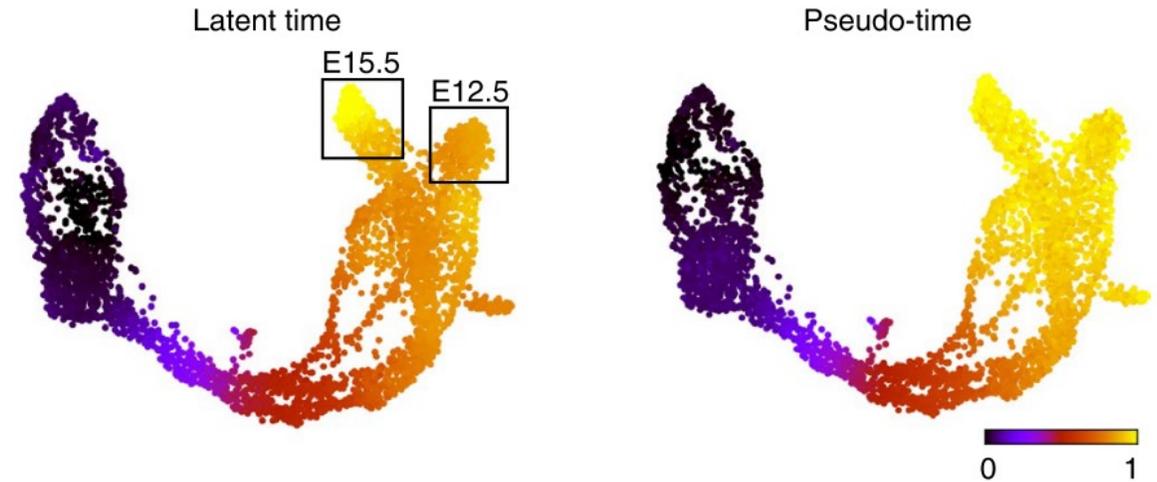
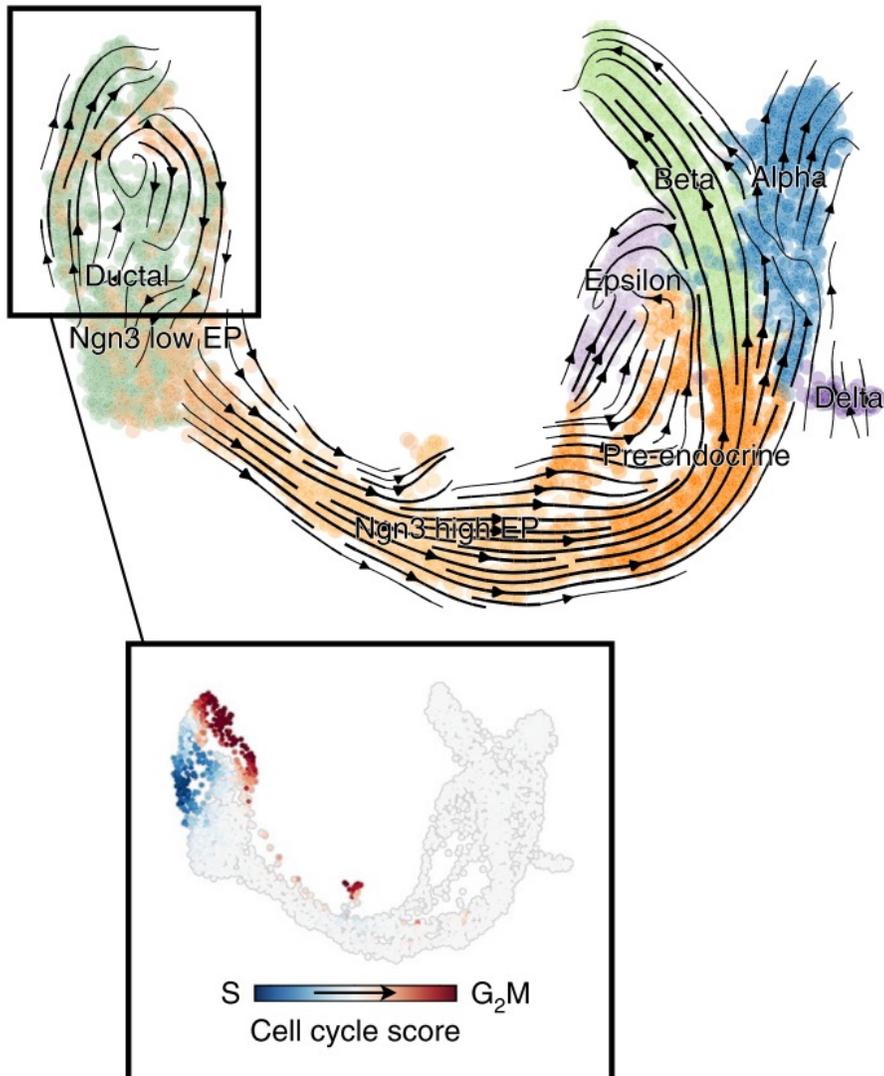
a likelihood for each gene

- explaining how well a cell is described by the learned spliced/unspliced phase trajectory.
- Aggregating over cells to obtain an overall gene likelihood

Results: Delineating cycling progenitors, commitment and fate transitions in endocrinogenesis

a

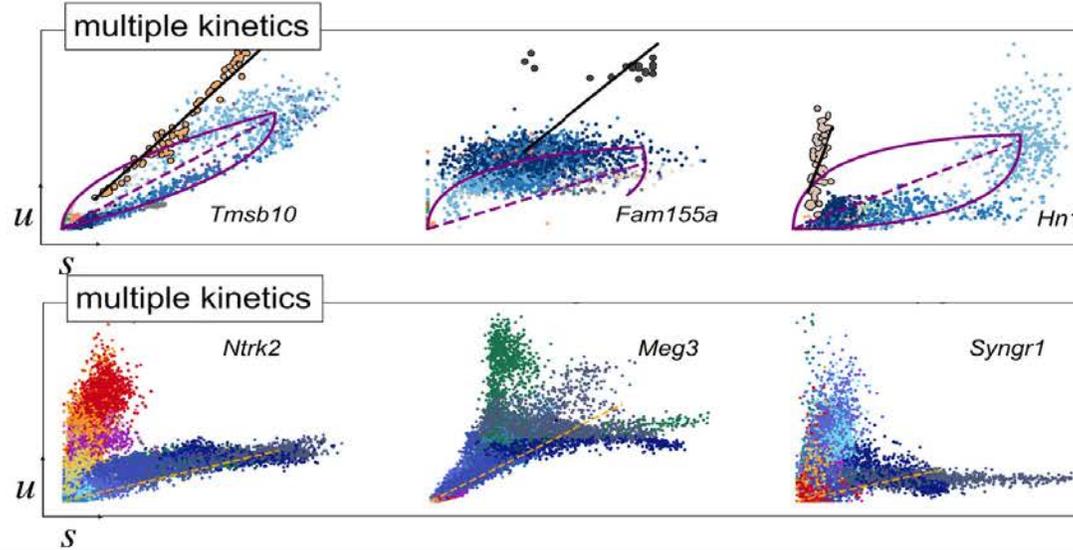
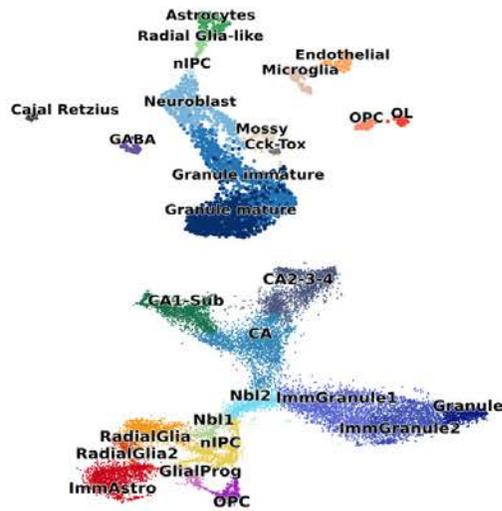
Dynamical model



- Capture cell cycle
- Predict real time better
 - α cells are produced earlier (before E12.5) then β cells (E12.5 – E15.5)

Potential pitfalls using current method

A Multiple kinetic regimes in Dentate Gyrus



Multiple kinetics

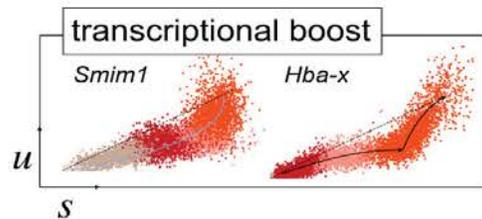
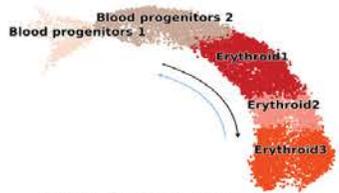
- different genes have different kinetics

Transcription boost

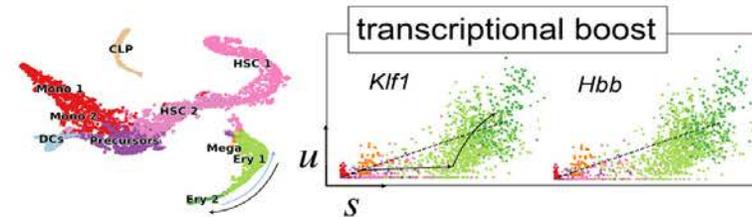
- for one gene it can have several different kinetics regime

B Transcriptional boost in erythroid maturation

Gastrulation erythroid maturation



Human bone marrow hemopoietic cells



Conclusion

- Cell-specific RNA velocity estimates provide a natural basis for quantitative modelling of cell fates.
- Identification of dynamics driving genes
- Capturing cell cycles, complex branchings and heterogenous subpopulation kinetics
- Inference of shared latent time which enables relating lineages

Limitation and outlook

- Assumes constant reaction rates; full-length protocols allow accounting for:
 - Time- and state-dependent rates;
 - Alternative splicing
- Incorporate time and spatial information.

Q&A

Thank you!